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(54) Title: DETECTION OF EXTRACELLULAR TUMOR-ASSOCIATED NUCLEIC ACID IN BLOOD PLASMA OR SERUM

(57) Abstract: This invention relates to detection of specific extracellular nucleic acid in plasma or serum fractions of human or animal blood associated with neoplastic, pre-malignant or proliferative disease. Specifically, the invention relates to detection of nucleic acid derived from mutant oncogenes or other tumor-associated DNA, and to those methods of detecting and monitoring extracellular mutant oncogenes or tumor-associated DNA found in the plasma or serum fraction of blood by using DNA extraction with or without enrichment for mutant DNA.

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DETECTION OF EXTRACELLULAR TUMOR-ASSOCIATED NUCLEIC ACID IN BLOOD PLASMA OR SERUM

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BACKGROUND OF THE INVENTION

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1. Field of the Invention

This invention relates to methods for detecting specific extracellular nucleic acid in plasma or serum fractions of human or animal blood associated with neoplastic, pre-malignant or proliferative disease. Specifically, the invention relates to detection of
15 nucleic acid derived from mutant oncogenes or other tumor-associated DNA, and to methods of detecting and monitoring extracellular mutant oncogenes or tumor-associated DNA found in the plasma or serum fraction of blood using DNA enrichment methods, wherein enrichment-based extraction methods are used prior to amplification and/or detection, or wherein enrichment for the nucleic acid of interest occurs during
20 amplification, in particular through use of a restriction endonuclease. In particular, the invention relates to the detection, identification, or inference of the existence of premalignant neoplasms or tissue in humans or other animals without cancer, wherein the neoplasm contains a mutation that is associated with the neoplasm, through detection of the mutated nucleic acid associated with the neoplasm in plasma or serum fractions.

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2. Description of the Related Art

Neoplastic disease, including most particularly that collection of diseases known as cancer, is a significant part of morbidity and mortality in adults in the developed
30 world, being surpassed only by cardiovascular disease as the primary cause of adult death. Although improvements in cancer treatment have increased survival times from diagnosis to death, success rates of cancer treatment are more closely related to early detection of neoplastic disease that enable aggressive treatment regimes to be instituted before either primary tumor expansion or metastatic growth can ensue. A particularly

favorable prognosis is achieved if premalignant tissue can be eradicated prior to progression to cancer.

Oncogenes are normal components of every human and animal cell, responsible for the production of a great number and variety of proteins that control cell proliferation, growth regulation, and cell death. Although well over one hundred
5 oncogenes have been described to date with nearly all identified at the deoxyribonucleic acid (DNA) sequence level. It is likely that a large number of oncogenes remain to be discovered.

Mutations in DNA occur as the result of inborn (inherited) genetic errors or
10 acquired (somatic) mutations, often as a result of environmental insults, and have long been recognized as playing a causative role in the development of neoplastic disease.

Within the last twenty years the sites of such mutations have been recognized to be within oncogenes, including tumor suppressor genes, and similar tumor-related gene regions including those with DNA microsatellite alterations and hypermethylated genes.
15 Mutation of these genes have been found to be an intrinsic and crucial component of premalignant and malignant growth in both animals and humans. Many malignant tumors or cell lines derived from them have been shown to contain one or more mutated oncogenes, and it is possible that every tumor contains at least one mutant oncogene.

Mutated oncogenes are therefore markers of malignant or premalignant
20 conditions. It is also known that other, non-oncogenic portions of the genome may be altered in the neoplastic state. Nucleic acid based assays can detect both oncogenic and non-oncogenic DNA, whether mutated or non-mutated.

In particular, nucleic acid amplification methods (*for example*, the polymerase chain reaction) allow the detection of small numbers of mutant molecules among a
25 background of normal ones. While alternate means of detecting small numbers of tumor cells (such as flow cytometry) have generally been limited to hematological malignancies (Dressler and Bartow, 1989, *Semin. Diag. Pathol.* 6: 55-82), nucleic acid amplification assays have proven both sensitive and specific in identifying malignant cells and for predicting prognosis following chemotherapy (Fey *et al.*, 1991, *Eur. J.*
30 *Cancer* 27: 89-94).

Various nucleic acid amplification strategies for detecting small numbers of mutant molecules in solid tumor tissue have been developed, particularly for the *ras* oncogene (Chen and Viola, 1991, *Anal. Biochem.* 195: 51-56; Kahn *et al.*, 1991, *Oncogene* 6: 1079-1083; Pellegata *et al.*, 1992, *Anticancer Res.* 12: 1731-1736; Stork *et al.*, 1991, *Oncogene* 6: 857-862). For example, one sensitive and specific method identifies mutant *ras* oncogene DNA on the basis of failure to cleave a restriction site at the crucial 12th codon (Kahn *et al.*, 1991, *ibid.*). Similar protocols can be applied to detect any mutated region of DNA in a neoplasm, allowing detection of other oncogene DNA or tumor-associated DNA. Since mutated DNA can be detected not only in the primary cancer but in both precursor lesions and metastatic sites (Dix *et al.*, 1995, *Diagn. Molec. Pathol.* 4: 261-265; Oudejans *et al.*, 1991, *Int. J. Cancer* 49: 875-879), nucleic acid amplification assays provide a means of detecting and monitoring cancer both early and late in the course of disease.

While direct analysis of neoplastic tissue is frequently difficult or impossible (such as in instances of occult, unrecognized disease), peripheral blood is easily accessible and amenable to nucleic acid-based assays such as those mentioned above. Many studies have used nucleic acid amplification assays to analyze the peripheral blood of patients with cancer in order to detect intracellular DNA extracted from circulating cancer cells in patients, including one study which detected the intracellular *ras* oncogene from circulating pancreatic cancer cells (Tada *et al.*, 1993, *Cancer Res.* 53: 2472-4). However, it must be emphasized that these studies attempt to use nucleic acid-based amplification assays to detect extracted intracellular DNA within circulating cancer cells. The assay is performed on the cellular fraction of the blood from patients having cancer using the cell pellet or cells within whole blood, and the serum or plasma fraction is conventionally ignored or discarded prior to analysis. Since such an approach requires the presence of metastatic circulating cancer cells (for non-hematologic tumors), it is of limited clinical use in patients with early cancers, and it is not useful in the detection of non-hematologic non-invasive neoplasms or pre-malignant states.

It was known in the prior art that small but significant amounts of normal DNA circulate in the blood of healthy people (Fedorov *et al.*, 1986, *Bull. Exp. Biol. Med.*

102: 1190-2; Leon *et al.*, 1977, *Cancer Res.* 37: 646-50), and this amount has been found to increase in cancer states (Shapiro *et al.*, 1983, *Cancer* 51: 2116-20; Stroun *et al.*, 1989, *Oncology* 46: 318-322). The prior art contains disclosure that mutant oncogene DNA could be detected in peripheral blood plasma or serum of cancer patients
5 (see, for example, Sorenson *et al.*, 1994, *Cancer Epidemiology, Biomarkers & Prevention* 3: 67-71; Vasioukhin *et al.*, 1994, *Br. J. Haematol.* 86: 774-9; Vasyukhin *et al.*, in Verna & Shamoo (eds), *Biotechnology Today*, Ares-Serono Symposia Publications, pp. 141-150). Mutant ras oncogenes have been demonstrated in plasma or serum using polymerase chain reaction. However, these reports have also been
10 generally limited to patients with advanced cancer or known neoplastic or proliferative disease.

We have recognized that nucleic acid amplification assays can detect tumor-associated extracellular mutated DNA, including oncogene DNA, in the plasma or serum fraction of blood of humans without clinically-diagnosed cancer or known disease (see
15 U.S. Serial No. 08/818,058, incorporated by reference), and that this can be accomplished in a clinically useful manner.

SUMMARY OF THE INVENTION

20 The present invention relates to detection of specific extracellular nucleic acid in plasma or serum in human or animals without cancer which are associated with neoplastic, preneoplastic or proliferative disease or conditions. Specifically, the invention relates to the detection of premalignant disease by the direct enrichment of mutated nucleic acid or tumor-associated nucleic acid found in the plasma or serum
25 fraction of blood with respect to wild-type or non-mutated nucleic acid either prior to or during amplification and detection of the nucleic acid, whereby the concentration of the mutant nucleic acid is increased or the mutant nucleic acid is isolated from the remaining non-mutated nucleic acid. The invention thus provides methods for detecting predictive risk markers for a variety of cancers, including colorectal, pancreatic, lung,
30 prostate, esophageal, gastric, breast, bladder, ovarian, cervical, liver, and kidney cancer,

and other malignancies and premalignant conditions carrying tumor-associated mutations in DNA, as well as methods for monitoring neoplastic disorders in humans and animals. The invention provides methods for detecting mutant oncogenes, including but not limited to mutated K-ras, APC, and P53 DNA in plasma or serum. Premalignant diseases or conditions include but are not limited to colorectal adenoma, cervical dysplasia, atypical squamous metaplasia of the lung, bronchial dysplasia, atypical hyperplasia of the breast, prostatic intraepithelial neoplasia, atypical endometrial hyperplasia, dysplastic nevi of the skin, and Barrett's esophagus.

The prior art provides instruction in identifying tumor-associated DNA in the plasma or serum fraction of blood of humans with known malignancy using DNA extraction methods, followed by amplification of the target DNA, followed by detection of the amplified target nucleic acid (*see, for example*, Sorenson *et al.*, 1993, American Association for Cancer Research Abstract #174; Vasyukhin *et al.*, 1994, *Biotechnology Today* (Verna & Shamoo, eds.), Ares-Serono Symposia Publications, pp.141-150; Vasioukhin *et al.*, 1994, *Br. J. Haematol.* 86: 774-9; Lefort *et al.*, 1995, American Association for Cancer Research Abstract #557; Nawroz *et al.*, 1996, *Nature Med.* 2:1035-7; Chen *et al.*, 1996, *Nature Med.* 2: 1033-5). However, the prior art does not apply these methods to non-hematologic premalignancy, or to patients without known disease. The methods disclosed herein, in contrast, allow detection of mutant DNA or tumor-associated DNA from the blood of humans without cancer or known disease by providing for the enrichment of mutated nucleic acid, wherein the mutated nucleic acid is concentrated and/or isolated from the remaining extracted nucleic acid prior to or independent of amplification of the target nucleic acid, and thereby provides methods which enable enhanced detection of the target nucleic acid or a fragment thereof. The methods disclosed herein further provide for the enrichment of mutated nucleic acid with respect to wild-type nucleic acid during amplification of the target nucleic acid, and thereby provide for methods which enable the enhanced detection of the target nucleic acid or a fragment thereof.

Extracellular DNA is known to circulate in the serum or plasma fraction of blood (Stroun *et al.*, 1987, *Eur. J. Cancer Clin. Oncol.* 23: 707-12; Stroun *et al.*, 1989,

Oncology 46: 318-322). The invention disclosed in co-owned and co-pending U.S. Serial No. 08/818,058 (incorporated by reference) taught methods for identifying mutated extracellular DNA for identification of premalignant lesions. These methods are useful for diagnosis and treatment of people who are at risk to develop malignancy or premalignancy. Appropriate therapy, including but not limited to, increased surveillance, surgical excision, chemotherapy and immunotherapy or chemoprevention therapies, as well as more innovative therapies (such as antisense oligonucleotide therapy directed at a mutated *KRAS* oncogene or vaccine therapy, for example), may be instituted based on detection of mutant or non-mutated tumor-associated nucleic acid in such patients.

Prior to the instant invention and co-owned and co-pending US serial No. 08/818,058 it was not known that extracellular mutant nucleic acid could be detected in the blood of humans without cancer. The present invention and co-owned and co-pending US serial No. 08/818,058 teach that detection of extracellular mutant nucleic acid in the blood of humans without cancer can be enhanced by enrichment of the mutated nucleic acid relative to wild-type nucleic acid wherein the mutated nucleic acid is concentrated and/or isolated from the remaining extracted nucleic acid in a manner performed either concurrently or sequentially with the extraction and prior to amplification and/or detection of the target nucleic acid or a fragment thereof. Although extracellular mutant tumor-associated DNA has been detected in patients with advanced malignancies (Sorenson *et al.*, 1993, *Id.*; Vasioukhin *et al.*, 1994, *Id.*; Nawroz *et al.*, 1996, *Id.*), it had been presumed in the prior art that only malignancies, and in particular large malignancies, produced enough extracellular DNA to be identified even with prior amplification of the DNA. Co-owned and co-pending U.S. Serial No. 08/818,058 (incorporated by reference) taught that mutated oncogenes and other tumor-associated nucleic acid was also detectable in blood serum or plasma of individuals with pre-malignant lesions, diseases, or conditions following amplification of nucleic acid sequences found in blood plasma or serum. In the present invention additional methods are taught which enable enrichment of mutated nucleic acid thereby providing enhanced detection of nucleic acid or a fragment thereof whereby detection is indicative or associated with the presence of premalignant tissue in the human.

The present invention provides methods for detecting the presence of extracellular nucleic acid in blood plasma or serum fractions, said nucleic acid being associated with a neoplastic, pre-malignant or proliferative disease state in an animal or a human without cancer. The invention provides methods for extracting and enriching
5 extracellular nucleic acid associated with a neoplastic, pre-malignant or proliferative disease state in an animal or a human prior to nucleic acid amplification or signal detection. The invention further provides for methods whereby mutated nucleic acid is enriched with respect to wild-type nucleic acid during or prior to an amplification step wherein a restriction endonuclease is used during or prior to the amplification step.
10 These methods of the invention are used for the detecting, monitoring, evaluating, or risk assessment of premalignant conditions, and in particular those conditions including but not limited to colorectal adenoma, cervical dysplasia, atypical squamous metaplasia of the lung, bronchial dysplasia, atypical hyperplasia of the breast, prostatic intraepithelial neoplasia, atypical endometrial hyperplasia, dysplastic nevi of the skin, and Barrett's
15 esophagus.

In a first aspect, the invention provides a method for detecting extracellular tumor-derived or tumor-associated mutated nucleic acid in a plasma or serum fraction of a blood sample from a human or animal without cancer, thereby providing a method for diagnosis, detection, monitoring, evaluation or treatment of a neoplastic or
20 proliferative disease in an animal or a human. The method provided by the invention comprises the steps of: first, purifying extracellular nucleic acid from blood plasma or serum to prepare a preparation of extracted nucleic acid containing a tumor-associated mutated nucleic acid or a fragment thereof; second, enriching for the mutated nucleic acid, either concurrent with or sequentially following the initial extraction step, wherein
25 the mutated nucleic acid or a fragment thereof is concentrated and/or isolated from the remaining extracted nucleic acid; third, amplifying the enriched mutated nucleic acid or a fragment thereof, or amplifying a signal corresponding to the enriched mutated nucleic acid or a fragment thereof; and fourth, detecting the product of the amplified mutated nucleic acid or a fragment thereof, or the amplified signal corresponding to the extracted
30 mutated nucleic acid or a fragment thereof, wherein the mutated nucleic acid or a

fragment thereof is associated with neoplastic, pre-malignant or proliferative disease.

In preferred embodiments of this aspect of the invention, the mutated nucleic acid is derived from nucleic acid encoding a mutated oncogene or other tumor-associated DNA, such as a DNA microsatellite alteration.

5 In another aspect, the invention provides a method for detecting extracellular tumor-derived or tumor-associated nucleic acid in a plasma or serum fraction of a blood sample from a human or animal without clinically-diagnosed cancer, thereby providing a method for detection, diagnosis, monitoring, evaluation, or treatment of a neoplastic or proliferative disease or premalignant conditions in an animal or a human. The method
10 provided by the invention comprises the steps of: first, purifying extracellular nucleic acid from plasma or serum to prepare a preparation of extracted nucleic acid containing a tumor-associated mutated nucleic acid or a fragment thereof; second, enriching the mutated nucleic acid relative to wild-type nucleic acid using an endonuclease prior to or during amplification; third, amplifying the enriched mutated acid or a fragment
15 thereof; and fourth, detecting the amplified fragment of the mutated nucleic acid or a signal corresponding to the amplified fragment of the mutated nucleic acid. In preferred embodiments of this aspect of the invention, the nucleic acid is derived from a nucleic acid encoding an oncogene or other tumor-associated DNA.

 Particularly preferred embodiments of the invention comprise detection of
20 nucleic acid sequences derived from or related to mutated p53, *K-ras*, and APC alleles.

 In preferred embodiments of the inventive methods, extracellular nucleic acid is extracted from blood plasma or serum using an extraction method including gelatin extraction; silica, glass bead, or diatom extraction; guanidine- or guanidinium-based extraction; chemical extraction methods; and size-exclusion and anion-exchange
25 chromatographic methods. In particularly preferred embodiments, the target DNA is extracted in an enriching manner, or extracted DNA is further enriched, using probe-specific hybridization wherein said hybridizing probes are immobilized to a substrate, wherein such substrate includes but is not limited to nylon and magnetic beads, from which contaminating species (nucleic acid and otherwise) can be removed using
30 methods (such as washing at defined stringencies of salt concentration and temperature)

known in the prior art, or wherein alternatively the target DNA of interest may be otherwise isolated from contaminating species including wild-type DNA, for example by application of a magnetic field or an electric field. In preferred embodiments, the extracted and enriched nucleic acid is amplified or signal amplified, wherein the
5 amplification method may include but is not limited to polymerase chain reaction, ligase chain reaction, boomerang DNA amplification, strand displacement amplification, strand displacement activation, cycling probe amplification, and branched DNA signal amplification. In preferred embodiments, DNA detection is performed using a detection method including gel electrophoresis; immunological detection methods; hybridization
10 using a specific, fluorescent-, radioisotope-, antigenic- or chromogenically-labeled probe; Southern blot analysis; electrochemiluminescence; reverse dot blot detection; and high-performance liquid chromatography.

In a preferred embodiment, the nucleic acid is extracted from blood serum or plasma by heating the serum or plasma, at a temperature from about 90°C to about
15 100°C, more preferably from about 95°C to about 100°C, for a time from about 1 minute to about 20 minutes, more preferably from about 5 minutes to about 15 minutes, and most preferably from about 5 minutes to about 10 minutes. Optionally, the blood plasma or serum can be frozen after boiling to a temperature of from about -20°C to about 0°C for at least about 5 minutes, more preferably 15 minutes and most preferably for at least
20 about 30 minutes. The boiled blood plasma or serum is used after cooling or, if frozen, after being thawed to a liquid.

The methods of the invention are provided for detecting tumor-associated extracellular nucleic acid, for example a mutated oncogene, in a human without clinically-diagnosed cancer, whereby detection is indicative of the presence of non-
25 hemopoietic cells or tissue containing tumor-associated nucleic acid within the human, the method comprising the steps of purifying extracellular nucleic acid from a plasma or serum fraction of a blood sample from the human to prepare extracted nucleic acid containing a tumor-associated nucleic acid, for example a mutated oncogene DNA, or a fragment thereof, and concurrently or sequentially; enriching for the tumor-associated
30 nucleic acid or a fragment thereof, wherein the tumor-associated nucleic acid or a

fragment thereof is concentrated and/or isolated from the remaining extracted nucleic acid; and amplifying or signal amplifying the tumor-associated nucleic acid or a fragment thereof; and detecting the product of the amplified tumor-associated nucleic acid or a fragment thereof or the amplified signal of the tumor-associated nucleic acid or a fragment thereof. The detected nucleic acid or a fragment thereof is then identified, *e.g.*, as comprising the mutated form of an oncogene associated with a neoplastic, pre-malignant or proliferative disease, wherein detection of the amplified product or the amplified signal or the tumor-associated nucleic acid or a fragment thereof is indicative of the presence of non-hemopoietic cells or tissue having a mutated oncogene or other tumor-associated nucleic acid in the human. In a preferred embodiment, the methods of the invention are used as an aid in the diagnosis in a human of a neoplastic, pre-malignant or proliferative disease. In another preferred embodiment, the method is used to determine a predictive risk factor for a neoplastic disease or disease progression in a human. Additionally, the methods of the invention are preferably used to determine disease prognosis in a human. In other preferred embodiments, the methods of the invention are used to determine the need for additional diagnostic tests, or for treatment.

Particularly preferred methods of the invention enable detection in blood of mutated nucleic acid, wherein the mutation is an acquired (somatic) mutation, as may be optionally shown by demonstrating the absence of the mutation in normal cells from the human having the acquired mutation, such as in normal leukocytes or other normal tissue. The invention thus provides a method for determining an acquired predictive risk factor for a non-hematologic disease in a human without clinically-diagnosed cancer, the method comprising the steps of purifying extracellular nucleic acid from blood or blood plasma or serum from a human without clinically-diagnosed cancer to prepare extracted nucleic acid containing a mutated DNA or a mutated DNA fragment, and concurrently or sequentially enriching for the mutated DNA or a fragment thereof, wherein the mutated DNA or a fragment thereof is concentrated and/or isolated from the remaining extracted nucleic acid, and thereafter amplifying the enriched mutated DNA or a fragment thereof, or alternatively amplifying a signal from the enriched mutated DNA or a fragment thereof, and then detecting the product of the amplified mutated DNA or

the product of its amplified fragment, or the amplified signal of the mutated DNA or the amplified signal of a fragment thereof, whereby said detection determines a predictive risk factor for a non-hematologic disease. Further, and optionally, by demonstrating the absence of the mutated DNA in normal cells of the human, such as normal leukocytes
5 or other normal tissue, in which it is therefore demonstrated that the mutation is not an inherited or inborn mutation, the predictive risk factor is shown to be an acquired predictive risk factor. In a preferred embodiment the mutated DNA includes but is not limited to either a mutated oncogene DNA, for example mutated K-ras, P53, or APC, or a DNA microsatellite alteration. In preferred embodiments the non-hematologic
10 disease is either colorectal adenoma, cervical dysplasia, atypical squamous metaplasia of the lung, bronchial dysplasia, atypical hyperplasia of the breast, prostatic intraepithelial neoplasia, atypical endometrial hyperplasia, dysplastic nevi of the skin, or Barrett's esophagus.

Also provided as embodiments of the methods of the invention are methods
15 additionally comprising the steps of determining the nucleic acid sequence of the nucleic acid fragment of extracellular nucleic acid in the extracted nucleic acid fraction that is associated with neoplastic or proliferative disease, wherein the nucleic acid sequence of the nucleic acid fragment comprises a mutated or variant allele of a nucleic acid associated with a neoplastic or proliferative disease.

20 In addition to the methods noted above, the invention provides methods for isolating extracellular tumor-derived or tumor-associated nucleic acid from a fraction of a blood sample comprising the plasma fraction or the serum fraction of the blood sample. In these embodiments the method comprises the steps of purifying extracellular nucleic acid from plasma or serum to prepare a preparation of extracted nucleic acid
25 using an extraction method; enriching the extracted nucleic acid fraction for the portion of the fraction that is associated with neoplastic, pre-malignant or proliferative disease; and cloning the DNA fragments comprising the enriched nucleic acid fraction that is associated with neoplastic, pre-malignant or proliferative disease. Also provided in this aspect of the invention are recombinant genetic constructs comprising a nucleic acid
30 fragment that is associated with a neoplastic, pre-malignant or proliferative disease or

condition that is prepared using the methods of the invention. Ribonucleic acid transcribed from the recombinant genetic constructs of the invention are also provided, as well as protein produced from translation of said RNA, and methods for using the translated proteins and peptides of the invention as epitopes for the production of antibodies and vaccines.

The invention also provides a method for concentrating and/or isolating any extracellular mutated DNA or a fragment thereof present in plasma or serum from a human without clinically-diagnosed cancer for which specific oligonucleotide hybridization primers are available, the method comprising the steps of purifying extracellular nucleic acid from blood of a human without clinically-diagnosed cancer to prepare nucleic acid containing a mutated DNA or a mutated DNA fragment, and concurrently or sequentially hybridizing an oligonucleotide to the mutated DNA or a mutated DNA fragment to produce a hybridized product of mutated DNA or a fragment thereof, and separating the hybridized product of mutated DNA or a fragment thereof from the remaining non-hybridized nucleic acid, thereby concentrating and/or isolating the mutated DNA or a fragment thereof. The invention thus provides a method for enriching for a mutated oncogene allele in plasma or serum, including but not limited to mutated K-ras, P53, or APC, and enriching for a DNA having a microsatellite alteration. In a preferred embodiment, the signal from the specific nucleic acid fragments comprising the enriched nucleic acid fraction is amplified using methods known to those with skill in the art, for example, branched DNA signal amplification, and combinations or variations thereof. In a preferred embodiment, detection of specific DNA fragments is performed using a detection method such as gel electrophoresis, immunological detection methods, nucleic acid hybridization using a specific, fluorescent- or chromogenically-labeled probe, Southern blot analysis, electrochemiluminescence, reverse dot blot detection, or high-performance liquid chromatography.

The invention further provides a method for detecting a mutation in the DNA present in blood or a blood fraction, *i.e.*, plasma or serum, of a human having a disease or premalignant condition in whom the presence of the disease or premalignant condition is unrecognized, wherein the mutation is an acquired mutation of known

association with the disease or premalignant condition, and wherein the disease or premalignant condition includes but is not limited to colorectal adenoma, cervical dysplasia, atypical squamous metaplasia of the lung, bronchial dysplasia, atypical hyperplasia of the breast, prostatic intraepithelial neoplasia, atypical endometrial hyperplasia, dysplastic nevi of the skin, or Barrett's esophagus. The method comprises the steps of purifying extracellular nucleic acid from blood or a blood fraction from a human in whom the presence of a disease or premalignant condition is unrecognized to prepare extracted nucleic acid containing a mutated DNA or a fragment thereof wherein the mutation is of known association with the disease or premalignant condition, and amplifying the mutated DNA or a fragment thereof, or alternatively amplifying a signal from the mutated DNA or a fragment thereof, and detecting the product of the amplified mutated DNA or a fragment thereof, or the amplified signal of the mutated DNA or a fragment thereof. In a particularly preferred embodiment, the method enables the evaluation of blood or a blood fraction from a human for the mutated oncogene to assist in the identification of the premalignant disease or premalignant condition.

It is therefore the object of this invention to detect or infer the presence of precancerous cells from non-hematologic premalignancies, within a human or animal body having a recognized neoplastic or proliferative disease or in those not previously diagnosed, by examining the plasma or serum fraction of blood for extracellular mutated oncogene DNA or tumor-derived or associated extracellular DNA, using a nucleic acid detection assay.

Another object of this invention is to enable extraction of extracellular tumor-associated nucleic acid from plasma or serum in an enriching manner, wherein the tumor-associated nucleic acid is concentrated and/or isolated from the remaining extracted nucleic acid.

An advantageous application of this invention is to identify, either quantitatively or qualitatively, acquired mutant oncogenes or acquired tumor-associated DNA in the blood plasma or serum of humans or animals as to classify such patients for their risk of neoplastic disease.

Another advantageous application of this invention is to identify, either quantitatively or qualitatively, mutant oncogenes or tumor-associated DNA in the blood plasma or serum of humans or animals who are receiving therapies, including but not limited to chemopreventive therapy and polypectomy, as a guide to whether adequate therapeutic effect has been achieved or whether additional or more advanced therapy is required, and to assess prognosis in these patients.

Another advantageous application of this invention is to identify, either quantitatively or qualitatively, mutant oncogenes or tumor-associated DNA in the blood plasma or serum of humans or animals as to determine the need for additional diagnostic testing, wherein additional diagnostic testing includes but is not limited to endoscopy, colonoscopy, sigmoidoscopy, bronchoscopy, radiologic imaging, radionucleotide scanning, ultrasonography, PET scanning, or further evaluation of organ or site specific bodily fluids or stool, wherein bodily fluid includes but is not limited to that collected by drainage, aspiration, direct sampling, and lavage.

Another advantageous application of this invention is to identify, either by detection or inference, the presence of premalignant neoplasms, conditions, or diseases, through detection of acquired mutant oncogenes or acquired tumor-associated DNA in the blood of humans or animals in whom the presence of the neoplasm, condition, or disease is unrecognized, wherein the acquired mutant DNA derives from premalignant growths such as dysplasias or adenomas, or from other cells bearing an acquired mutated DNA of known association with a premalignant disease or condition, wherein said diseases and conditions include but are not limited to colorectal adenoma, cervical dysplasia, atypical squamous metaplasia of the lung, bronchial dysplasia, atypical hyperplasia of the breast, prostatic intraepithelial neoplasia, atypical endometrial hyperplasia, dysplastic nevi of the skin, and Barrett's esophagus. In addition, the invention advantageously provides a panel of several oncogene assays that can distinguish malignant from premalignant conditions, or assist in medical monitoring to detect transformation of the growth to an outright malignancy, or to detect regression.

Furthermore, the invention advantageously provides a means to define risk of malignancy in a human wherein the risk was previously unrecognized. Thus, the

invention provides methods for early identification of neoplasms and premalignant conditions of the colon, rectum, pancreas, lung, breast, bladder, ovary, cervix, endometrium, liver, prostate, and stomach and of other neoplasms or premalignant conditions carrying tumor-related mutations of DNA, and methods for monitoring
5 neoplastic disorders in humans and other animals.

Advantageous application of the invention is provided as example but not limitation to include detection of mutated K-ras allele in blood from a human with adenoma; detection of mutated P53 allele in blood from a human with adenoma; detection of mutated APC allele in blood from a human with a colorectal cancer;
10 detection of mutated K-ras allele in blood from a human with a proliferative disease; and detection of mutated K-ras allele in blood from a human with an in-situ (non-invasive or pre-invasive) carcinoma.

Thus, the invention provides a method of screening both healthy individuals and individuals at risk for cancer and premalignant conditions.

15 Another advantageous application of this invention is to identify, either quantitatively or qualitatively, one or more of multiple mutant oncogenes or tumor-associated DNAs in the blood plasma or serum of humans or animals without clinically-diagnosed cancer or not known to have cancer by use of a panel of oligonucleotide primers each specific for one of multiple differing oncogene alleles
20 detected by the panel, wherein a particularly preferred application employs a nucleic acid enrichment method, and wherein the oncogene alleles are detected concurrently or sequentially. Additionally, said panel enables the inference of the presence of specific tumor types based upon mutated oncogenes or other tumor-associated mutated nucleic acid detected by the panel. Additionally, said panel enhances identification of humans
25 at risk for neoplastic disease. Additionally, said panel enhances identification of humans having premalignant or malignant tissue. A particularly preferred advantageous application of this invention identifies one or more of either K-ras, P53, and/or APC mutated alleles in blood from a human without clinically-diagnosed cancer by use of a panel of primers specific for at least one or more of either K-ras, P53, or APC mutated
30 alleles, whereby a human at risk for a gastrointestinal neoplastic disease is identified.

The invention thereby provides for methods of detecting an acquired mutated APC allele or a fragment thereof in blood or a blood fraction.

Specific preferred embodiments of the present invention will become evident from the following more detailed description of certain preferred embodiments and the
5 claims.

DETAILED DESCRIPTION OF THE INVENTION

This invention provides methods for detecting or inferring the presence of precancerous cells in a human or animal. The method comprises preparation of nucleic
10 acid from plasma or serum wherein the extraction of nucleic acid may be performed with or without DNA precipitation or other separation of soluble nucleic acid from the plasma or serum, and further performed using a method of enrichment for mutated nucleic acid, wherein a mutated nucleic acid is concentrated and/or isolated, in either case followed by detection of mutated nucleic acid preferentially following amplification of the
15 mutated nucleic acid or its signal. Enrichment of the mutated nucleic acid may occur prior to or during amplification by methods provided, including methods using an endonuclease or by methods involving hybridization of the mutated nucleic acid. Embodiments of the invention which provide for enrichment of mutated DNA from blood or blood fractions offer the beneficial advantage of enhancing detection of the
20 mutated DNA. Direct sequencing of mutant nucleic acid is further made possible so that a broad range of mutated oncogenes including tumor suppressor genes, translocated genes, hypermethylated genes, microsatellite alterations in DNA, and non-coding DNA related to the development of malignancy may be identified. Embodiments of the invention provided that exhibit the absence of DNA precipitation methods from the
25 invention serves several beneficial functions. First, such methods reduce the manipulation required for each sample, thus decreasing time and cost in preparation. Second, such decrease in manipulation reduces the risk of contamination of one specimen with another or with exogenous DNA. Third, lack of precipitation steps decrease losses attendant with specimen handling and extraction, thus reducing the
30 sample volume required and potentially increasing the yield of DNA analyzed in each

specimen. Fourth, such methods increase the ease with which the entire analytical process may be automated, further reducing costs and turn-around time. Fifth, by reducing the variability in yield inherent in a precipitation process, such methods enable meaningful quantification of mutant nucleic acid, potentially providing prognostic information related to tumor burden.

The invention further provides a number of methods of identifying mutations without prior knowledge of their location to be employed. This significantly increases the utility of detecting mutated DNA in plasma or serum, since genes such as *APC* and p53 exhibit point mutations throughout their coding regions (Hollstein *et al.*, 1994, *Nucleic Acids Res.* 22: 3551-5). This is a significant improvement over the prior art, which requires prior knowledge of the location of a mutation or restriction of a search for mutants to the few most frequently mutated loci ("hot spots"; *see* Sorenson *et al.*, 1993, *Id.*; Vasioukhin *et al.*, 1994, *Id.*; Nawroz *et al.*, 1996, *Id.*; Sorenson *et al.*, 1994, *Cancer Epidemiology, Biomarkers & Prevention* 13: 67-71). Thus, a larger variety of malignant and premalignant conditions may be assessed (since different tumor types seem to carry particular mutations not found in other tumor types) and they may be detected for more thoroughly. It is to be noted and understood that the methods of DNA preparation and enrichment described herein are furthermore applicable to the previously described aspects of the invention utilizing amplification as well as methods of detection without amplification.

The invention provides methods for detection of an acquired (somatic) DNA mutation in the blood of a human without clinically-diagnosed cancer, wherein normal cells from the human lack the mutation; thereby the invention provides for a method of determining acquired predictive risk factors for non-hematologic acquired diseases and conditions.

Moreover, the assays and methods of the invention can be performed qualitatively, whereby the amount of the nucleic acid product produced is at least sufficient for efficient detection of the product, or quantitatively, whereby the amount of the nucleic acid product produced is measured with reference to a standard useful in determining the significance of the amount of produced nucleic acid (*for example*,

wherein the amount of nucleic acid product is related to a disease state or risk of developing a disease state).

Specifically, the invention provides methods for detecting nucleic acid in plasma or serum of a human or animal without clinically-diagnosed cancer wherein the nucleic acid is associated with the existence of pre-malignant cells or tissues in the human or animal, thereby providing a sensitive predictive risk factor for neoplastic disease and premalignant conditions, wherein such diseases and conditions include but are not limited to colorectal adenoma, cervical dysplasia, atypical squamous metaplasia of the lung, bronchial dysplasia, atypical hyperplasia of the breast, prostatic intraepithelial neoplasia, atypical endometrial hyperplasia, dysplastic nevi of the skin, and Barrett's esophagus.

The invention further provides methods for identification of humans having unrecognized disease who might therefore benefit from additional diagnostic testing, including but not limited to colonoscopy, sigmoidoscopy, endoscopy, bronchoscopy, radiologic imaging including CT scans, spiral CT scans, MRI, contrast studies, and plain films, ultrasonography, radionucleotide imaging, PET scanning, and evaluation of organ specific bodily fluid, stool, or lavage fluid, wherein bodily fluid includes but is not limited to that collected by drainage, aspiration, direct sampling, and lavage.

A General Overview of the Inventive Methods

In the practice of the invention blood is drawn by standard methods into a collection tube, preferably comprising siliconized glass, either without anticoagulant for preparation of serum or with EDTA, heparin, or similar anticoagulants, most preferably EDTA, for preparation of plasma. Plasma may optionally be subsequently converted to serum by incubation of the anticoagulated plasma with an equal volume of 0.025 molar calcium chloride at 37°C for a brief period, most preferably for 1-3 minutes, until clotting takes place. The clot may then be pelleted by a brief centrifugation (1-10 seconds) at 1,000x g or greater, and the deproteinized plasma removed to another tube. Preferably, the volume of plasma used is 5 L mixed with 5 L of calcium chloride, then diluted to 100 L with water. Alternatively, the centrifugation may be omitted. The

serum or plasma may be utilized directly for identification of mutant DNA. In one preferred embodiment, 10 L of serum or the prepared plasma is heated to a temperature greater than 90°C, most preferably greater than 94°C, for several minutes, most preferably 10. This heated substrate may then be cooled to below room temperature for a period of several minutes, or may be used directly in subsequent steps of the invention. In either instance an optional step of brief centrifugation at 1,000x g or greater may be performed to pellet any precipitate. Alternatively, heating may take place by placing a volume of sample, preferably 5 L, in a tube or microtiter well under mineral oil and heating in a microwave for greater than 3 minutes, more preferably 3-10 minutes (as described in Sanford *et al.*, 1997, *Biotechniques* 23: 890-2). In most preferred embodiments, nucleic acid is extracted from plasma or serum as an initial step of the invention. The extraction step may be performed either prior to or concurrent with the enrichment step.

15 Gelatin extraction method:

In a preferred embodiment, DNA is co-precipitated from plasma or serum with gelatin by a method modified from that of Fournie *et al.* (1986, *Anal. Biochem.* 158: 250-256).

A stock 5% (w/v) gelatin solution is prepared by mixing 1 gram gelatin (G8-500, Fisher, Pittsburgh, PA) with 20 mL sterile, double-distilled water, autoclaving for 30 minutes, and filtering through a 0.2 micron filter. The resultant solution is sequentially frozen in a dry ice/ethanol bath and thawed at room temperature for a total of five cycles.

A working 0.3% gelatin solution is prepared by heating the stock solution to 60°C and mixing 600 L of 5% gelatin with 25 L of 1 M Tris-HCl (pH 8.0) and 9.4 mL of sterile, double-distilled water.

25 Plasma or serum (160 L) is mixed with 12.8 L of 0.5 M EDTA and 467 L of sterile, double-distilled water, then emulsified for 3 minutes with 320 L of phenol or phenol:chloroform:isoamyl alcohol (25:24:1 ratio). The solution is centrifuged at 14,000 x g for 10 minutes, and 570 L of the aqueous layer is removed to a clean tube. DNA is precipitated by addition of 142 L of the 0.3% gelatin working solution and of 500 L of cold absolute ethanol, followed by incubation at -20°C for 1-2 hours. Extracellular

DNA may be precipitated within less than 1h of incubation at -20°C , and a very short incubation may be preferable in some circumstances. Alternatively, longer incubation at -20°C for 1-2 hours insures the precipitation of most DNA. The sample is centrifuged at $14,000 \times g$ at $4-6^{\circ}\text{C}$ for 15 minutes, washed once with cold 70% ethanol, and dried in
5 a 60°C heat block for 10 minutes. DNA is then recovered by the addition of 35 to 70 L of sterile, double-distilled water preheated to 60°C .

Glass bead, silica particle, or diatom extraction method.

As an alternative rapid method of extracting extracellular DNA from plasma or
10 serum, glass beads, silica particles, or diatoms may be used, as in the method or adaptation of Boom *et al.* (Boom *et al.*, 1991, *J. Clin. Microbiol.* 29: 1804-1811; Boom *et al.*, 1989, *J. Clin. Microbiol.* 28: 495-503). Size fractionated silica particles are prepared by suspending 60 grams of silicon dioxide (SiO_2 , Sigma Chemical Co., St. Louis, MO) in 500 mL of demineralized sterile double-distilled water. The suspension
15 is then settled for 24 hours at room temperature. Four-hundred thirty (430) mL of supernatant is removed by suction and the particles are resuspended in demineralized, sterile double-distilled water added to a final volume of 500 mL. After an additional 5 hours of settlement, 440 mL of the supernatant is removed by suction, and 600 L of HCl (32% wt/vol) is added to adjust the suspension to a pH 2. The suspension is aliquotted
20 and stored in the dark.

Lysis buffer is prepared by dissolving 120 grams of guanidine thiocyanate (GuSCN, Fluka Chemical, Buchs, Switzerland) into 100 mL of 0.1 M Tris hydrochloride (Tris-HCl) (pH 6.4), and 22 mL of 0.2 M EDTA, adjusted to pH 8.0 with NaOH, and 2.6 grams of Triton X-100 (Packard Instrument Co., Downers Grove, IL). The solution is
25 then homogenized.

Washing buffer is prepared by dissolving 120 grams of guanidine thiocyanate (GuSCN) into 100 mL of 0.1 M Tris-HCl (pH 6.4).

50 L of plasma or serum are mixed with 40 L of silica suspension prepared as above, and with 900 L of lysis buffer, prepared as above, using an Eppendorf 5432 mixer
30 over 10 minutes at room temperature. The mixture is then centrifuged at $12,000 \times g$ for

1 minute and the supernatant aspirated and discarded. The silica-DNA pellet is then washed twice with 450 μ L of washing buffer, prepared as above. The pellet is then washed twice with 1 mL of 70% (vol/vol) ethanol. The pellet is then given a final wash with 1 mL of acetone and dried on a heat block at 56°C for ten minutes. The sample is
5 eluted for ten minutes at 56°C with a TE buffer consisting of 10 mM Tris-HCl, 1 mM EDTA (pH 8.0) with or without Proteinase K (100 ng/ml) as described by Boom *et al.*, *ibid.* Following elution, the sample is then centrifuged at 12,000 x g for three minutes, and the DNA-containing supernatant recovered (Boom *et al.*, 1991, *ibid.*; Boom *et al.*, 1989, *ibid.*; Cheung *et al.*, 1994, *J. Clin. Microbiol.* 32: 2593-2597).

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Acid guanidinium thiocyanate-phenol-chloroform extraction method.

As an alternative method, extracellular DNA may be extracted from plasma or serum in one step using variations of the acid guanidinium thiocyanate-phenol-chloroform extraction method. For example, extracellular DNA may be extracted from
15 plasma or serum using TRI reagent, a monophasic guanidine-thiocyanate-phenol solution, as described by Chomczynski (1993, *Biotechniques* 15: 532-534). Plasma or serum (1 mL) is processed using 5-10 mL of TRI Reagent (commercially available as TRI Reagent, Molecular Research Center, Cincinnati, OH, and from other sources) according to manufacturer's directions. DNA is precipitated from the interphase with
20 ethanol.

Other commercially available kits known to the art may be utilized to extract nucleic acid from plasma or serum providing purified DNA for use in the invention's first step, including Qiagen columns (QIAamp blood kit, Qiagen, Basel, Switzerland),
25 Boehringer Mannheim's High-Pure Viral Nucleic Acid kit, and other commercial kits for extraction of nucleic acid from plasma or serum as described herein, which are provided as example and not as limitation. Such kits may be used according to manufacturer's directions.

30

Enrichment methods

Extracellular nucleic acid can be enriched from plasma, serum or whole blood using hybridization methods specific for particular species on extracellular nucleic acid.

For example, nucleic acid derived from mutant K-ras oncogene DNA can be enriched
5 from whole blood, serum or plasma using specific oligonucleotides for hybridization.

Such oligonucleotides, generally ranging in size from about 12 to about 15 nucleotides or longer, are advantageously centered around the mutated nucleotide of interest to provide the greatest degree of discrimination between mutant and wildtype alleles. In particular, for the K-ras oncogene, the nucleotides of interest are the first and second
10 positions of codons 12, 13 and 61. For K-ras, codon 12, first position, exemplary oligonucleotides have the sequence:

GTTGGAGCTCGTGGCGTAG (SEQ ID No.: 1)

GTTGGAGCTTGTGGCGTAG (SEQ ID No.: 2)

and

15 GTTGGAGCTAGTGGCGTAG (SEQ ID No.: 3),

where the underlined nucleotide in each oligonucleotide is mutated from the wildtype.

In another example, many mutations associated with the development of malignancy have been identified in the tumor suppressor gene p53 from a number of different tumors and tumor types. In colorectal carcinoma, for example, the most
20 common mutation involves codon 175. For this mutation, exemplary enrichment oligonucleotides include the following:

CCATGAGCACTGCTCAG (SEQ ID No.: 4)

CCATGAGCTCTGCTCAG (SEQ ID No.: 5)

and

25 CCATGAGCCCTGCTCAG (SEQ ID No.: 6),

where the underlined nucleotide in each oligonucleotide is mutated from the wildtype.

It will be apparent to one of ordinary skill in the art that any DNA point mutation can be enriched using oligonucleotides specific for the mutation and prepared to encompass the mutated site.

Additionally, enrichment oligonucleotides can be prepared to enrich for a particular gene such as an oncogene regardless of the presence of a mutation by preparing an oligonucleotide specific for the gene that does not encompass a site known to be involved in mutation related to the development of malignancy. Using such enrichment strategies, mutated nucleic acid is subsequently identified in the detection step of the methods of the invention, using mutation-specific oligonucleotides in hybridization assays, for example, or during a selective amplification step. For K-ras, enrichment oligonucleotides for nucleic acids encoding regions other than those containing codons 12, 13 or 61 are used to enrich a plasma, serum or whole blood sample for K-ras encoding nucleic acid sequences, both mutant and wildtype. Differentiation between mutant and wildtype fragment is then accomplished using detection methods such as allele-specific oligonucleotide hybridization assays.

Advantageously, more than one enrichment oligonucleotide species can be used simultaneously or sequentially. A panel of such enrichment oligonucleotides could include oligonucleotides specific for different mutated sites in a particular oncogene, or several mutated oncogenes in a multiplex enrichment assay. Differential detection methods, such as hybridization with distinguishable detectable labels, are then used to detect the different mutated nucleic acids resulting from the enrichment step of the methods of the invention.

Additional enrichment oligonucleotides include those oligonucleotides disclosed in co-owned and co-pending U.S. Serial No. 08/818,058, the disclosure of which is explicitly incorporated herein.

Additional nucleic acid extraction methods

Alternate means of purification which may be used to obtain DNA from serum or plasma, including selective retention on a size exclusion column or similar matrix, salting-out method, and other guanidinium thiocyanate extraction methods known in the art, including but not limited to all methods described in co-owned and co-pending U.S. serial No. 08/818,058.

Alternatively, the plasma or serum may be prepared as described above, and one or several regions of DNA of interest may be enriched. This may be performed by hybridization of nucleic acid in the heated plasma or serum solution or of extracted nucleic acid against a manufactured strand of nucleic acid that is complementary to the sequence of interest (an affinity capture approach; *see Seeger et al.*, 1997, *Biotechniques* 23: 512-7). Such "capturing" nucleic acids may consist of oligonucleotides, DNA or RNA constructs, or peptide nucleic acids. The capture of the target DNA serves as a means of enriching the target at the expense of other, non-hybridizing DNA that would otherwise compete with the target DNA during subsequent detection. The enrichment procedure occurs by hybridization at a low temperature under nonstringent conditions (*see generally*, Sambrook *et al.*, 1989, Molecular Cloning: A Laboratory Manual. (2nd ed.) Cold Spring Harbor Laboratory Press, New York), followed by washing of the target-capture complex to remove unbound, competing DNA. The capture nucleic acid may be bound to a surface, such as the wall of a tube or a nylon or other membrane, or it may be bound to glass, magnetic or other beads that are incubated with the serum or plasma (Gelsthorpe *et al.*, 1997, *Biotechniques* 22:1080-2; Rudi *et al.*, 1997, *Biotechniques* 22: 506-11), thereby enabling separation of the mutated nucleic acid alleles from wild-type nucleic acid alleles, for example but not limitation, by using a magnetic or electric field, or by other methods well known to the art. A single region of interest may be examined in each sample, or several regions may be enriched for. By judicious choice of capturing nucleic acids, more than one target DNA may be captured simultaneously. It will be understood by one skilled in the art that this multiplexing approach will speed analysis of samples, and may be performed in a relatively tumor-specific fashion, *i.e.*, with a set of capture probes representing genes that are mutated frequently in one type of tumor and infrequently in others. As an example, the genes *TP53*, *KRAS*, *DCC* and *APC* are often mutated in colorectal cancers and colorectal adenomas (precancerous tumors), while *VHL* and *WT1* are more often mutated in renal cell carcinomas. Thus, the identification of mutations in a series of genes or tumor-associated DNA may, beyond identifying the presence of a malignancy or premalignancy in a patient, serve to identify the particular type of neoplasia by the

particular set of genes or tumor-associated DNAs mutated or predict for the location of the neoplasia. Additionally, methods involving enrichment of the target nucleic acid may be performed on unprocessed plasma, serum or whole blood.

Alternatively, other means of enriching for the nucleic acid of interest in a
5 plasma or serum sample may be used. For example, antibodies directed against any portion of the target DNA may serve to capture it, with subsequent identification of the presence or absence of mutations. However, several antibodies may be needed to capture the entire gene or DNA of interest if this is desired, since we and others have found that DNA in plasma or serum often circulates in relatively small pieces, on the order of
10 several hundred base pairs in length. Such capturing antibodies may be raised in animals by immunization with the target DNA or fragments thereof, or may be purified from naturally occurring anti-DNA antibodies found in humans with rheumatologic conditions. As explained above, the capturing antibodies may be bound to tubes or wells as in an ELISA, or may be bound to tubing through which a sample travels prior to
15 further analysis, as by gas chromatography/mass spectroscopy or high performance liquid chromatography or bound to beads. These means and methods are provided by way of example and are not intended to be limiting.

Following the preparation of plasma or serum as described above, and preferably following an amplification step, a detection step for mutations in oncogenes or tumor-associated DNA is performed. It is the detection of mutants that indicates the presence
20 of one or more mutant-bearing cells within the patient. Moreover, as explained above, the pattern of mutants may be relatively specific for different types of proliferative diseases. It bears stressing that this invention, comprising either precipitation of extracellular DNA from serum or plasma or preparation of serum, plasma or whole
25 blood for analysis without precipitation of DNA followed by detection of mutations (including, but not limited to, point mutation, insertion, deletion and translocation) may be applied to detection of malignant neoplasms and detection of precursor, premalignant conditions.

A mutated DNA detected in blood plasma or serum of a human may be further
30 characterized as being an acquired (somatic) mutation by demonstrating the absence of

the mutation in normal cells of the human using methods known in the art consisting of extraction of the DNA from normal cells, and amplifying for the mutated DNA of interest in a manner as to enable detection, wherein an absence of the mutated DNA in normal cells from the human indicates the mutation detected within blood plasma or serum DNA to be an acquired mutation, and not an inherited or inborn mutation.

Detection of DNA sequence mutants may proceed by any of a number of methods known to those skilled in the art (Kilger *et al.*, 1997, *Nucleic Acids Res.* 25: 2032-4). The sequence may be detected directly by nucleic acid sequencing methods such as cycle sequencing (Sarkar *et al.*, 1995, *Nucleic Acids Res.* 23: 1269-70) or direct dideoxynucleotide sequencing, in which some or all of the enriched DNA of interest that has been harvested from plasma or serum is used as a template for sequencing reactions. An oligonucleotide primer or set of primers specific to the gene or DNA of interest is used in standard sequencing reactions.

Other methods of DNA sequencing, such as sequencing by hybridization, sequencing using a "chip" containing many oligonucleotides for hybridization (as, for example, those produced by Affymetrix Corp.; Ramsay *et al.*, 1998, *Nature Biotechnology* 16: 40-44; Marshall *et al.*, 1998, *Nature Biotechnology* 16: 27-31), sequencing by HPLC (DeDionisio *et al.*, 1996, *J. Chromatogr A* 735: 191-208), and modifications of DNA sequencing strategies such as multiplex allele-specific diagnostic assay (MASDA; Shuber *et al.*, 1997, *Hum. Molec. Genet.* 6: 337-47), dideoxy fingerprinting (Sarkar *et al.*, 1992, *Genomics* 13: 441-3; Blaszyk *et al.*, 1995, *Biotechniques* 18: 256-60; Martincic *et al.*, 1996, *Oncogene* 13: 2039-44), and fluorogenic probe-based PCR methods (such as Taqman; Perkin-Elmer Corp.; Heid *et al.*, 1996, *Genome Res.* 6: 986-94) and cleavase-based methods may be used. Alternatively, approaches that detect specific mutations of interest such as allele-specific amplification or restriction digest methods such as CARD (as disclosed in co-owned and co-pending U.S. Serial No. 08/818,058, incorporated by reference) may be used singly or in combination to identify extracellular mutant DNA (Zafiroopoulos *et al.*, 1997, *Biotechniques* 23:1104-1109). It will be clear to one skilled in the art that a variety of suitable methods for determining mutations at the DNA sequence level would suffice

for the practice of this step of the inventive methods, and the methods mentioned herein are not intended to be comprehensive or limiting, but merely to serve as examples.

Methods that detect mutations in DNA without precisely identifying the mutated base or bases are also able to be used in this invention, inclusive of those methods of
5 detection previously described in the application. Single strand conformation polymorphism (SSCP) analysis, for example, can identify variations from wildtype in a region of DNA without precisely defining the mutated base(s)(Orita *et al.*, 1989, *Proc. Natl. Acad. Sci. USA* 86: 2766-70; Suzuki *et al.*, 1990, *Oncogene* 5:1037-43). Similarly, heteroduplex analysis (Glavac *et al.*, 1995, *Hum. Mutat.* 6(4):281-7), denaturing gradient
10 gel electrophoresis (Pellegata *et al.*, 1992, *Anticancer Res.* 12:1731-1736) and mismatch cleavage assays can identify a patient carrying a DNA mutation by analysis of extracted or unextracted plasma or serum DNA. All of these procedures require cleavage of the sample DNA with restriction endonucleases, or alternatively amplification by PCR or a similar amplification technique, to obtain uniform sized DNA fragments for gel
15 electrophoresis analysis.

Detection of the target mutations can also be accomplished by means of signal amplification techniques. For example, the branched DNA assay (Chiron) uses a specific DNA probe to a target DNA (in this case, any of the oncogenes or tumor-associated DNAs of interest) to identify the presence of the target (Urdea *et al.*, 1993, *AIDS* 7: S11-
20 4). The signal is amplified by means of modifications made to the probe which allow many fluorescent detector DNA molecules to hybridize to a target nucleic acid. Similarly, oligonucleotide ligation assays may be used to amplify and detect the mutation of interest from amplified or directly processed serum or plasma DNA (Benson *et al.*, 1996, *Thromb. Res.* 83: 87-96). Any signal amplification method may be used in
25 the invention.

For the purposes of this invention, tumor-derived or associated DNA includes but is not limited to DNA related to mutated oncogenes or other mutated DNA, a partial list of which includes H-ras, K-ras, N-ras, c-myc, her-2/neu, bcr-abl, fms, src, fos, sis, jun, bcl-2, bcl-2/IgH, or VHL (Von Hippel-Lindau gene), and DNA microsatellite

alterations; DNA related to tumor suppressor genes, a partial list of which includes p53, RB, MCC, APC, DCC, NF1, WT1; and hypermethylated DNA.

It will be understood that methods of detection and/or enrichment for extracellular mutated nucleic acid from blood or blood fractions as described herein may
5 similarly be applied as methods for detection and/or enrichment of extracellular mutated nucleic acid present in other bodily fluids, including particularly ascitic fluid, pleural effusions, pericardial effusions, sputum and bronchial secretions, breast fluid including secretion from the ducts and nipple of the breast, gastric secretions, and fluid aspirated or drained from cystic or semi-cystic tissues, wherein evaluation of the bodily fluid
10 offers distinct diagnostic advantage in humans in whom the presence of a disease is unknown.

Methods of enrichment using an endonuclease.

15 Enrichment of the target or mutated nucleic acid, whereby the concentration of the target nucleic acid is increased with respect to wild-type or non-mutated nucleic acid, may be accomplished by using an endonuclease either before or during an amplification step, as disclosed in co-owned and co-pending U.S. serial No. 08/818,058, the disclosure of which is explicitly incorporated herein. In preferred embodiments, the endonuclease
20 may be *Bst*NI, *Hin*P I, or *Msp* I.

Therapeutic Applications

The extraction of extracellular DNA from plasma or serum permits further analysis or other manipulation of that DNA, from which further clinical utility is
25 realized. In this optional step of the invention, extracellular nucleic acid is analyzed to define the characteristics or composition of the tumor or pre-neoplastic lesion from which the nucleic acid originates. Any of several methods may be used, dependent upon the desired information, including nucleic acid sequencing, spectroscopy including proton NMR spectroscopy, biochemical analysis, and immunologic analysis. In the
30 preferred embodiment, such nucleic acid is cloned into a plasmid vector, for example the

pGEM-T vector plasmid (Promega, Madison, WI) and sequenced using a commercial kit such as Sequenase 2.0 (USB, Cleveland, OH). Analysis to define the characteristics or composition of the extracellular nucleic acid, and thus the characteristics of the originating tissue, affords a wide array of clinical utility, including the description, characterization, or classification of the tumor, whether known or occult, by tissue of origin, by type (such as premalignant or malignant), phenotype, or genotype, or by description or characterization of tumor behavior, physiology and biochemistry. This information is useful to characterize the lesion for tumor invasiveness, propensity to metastasize, and sensitivity or resistance to various therapies, thereby allowing the prediction of response to either ongoing or planned therapy and, further, allowing evaluation of prognosis. Comparison of the characteristics of extracellular nucleic acid to previous biopsy or surgical specimens permits further evaluation of tumor heterogeneity or similarity in comparison to that specimen, and thus evaluation of tumor recurrence or progression.

Following extraction of extracellular DNA from plasma or serum, complementary ribonucleic acid (RNA) may be transcribed or manufactured from the DNA. In a preferred embodiment, transcription of RNA is performed by employing a primer with an RNA polymerase promoter region joined to the standard primer sequence for the DNA of interest. RNA complementary to the DNA is then transcribed from the attached promoter region. In an alternative method, extracellular nucleic acid is cloned into an expression vector, and RNA complementary to the DNA is transcribed. Furthermore, as an optional preferred embodiment, the complementary RNA is used in an *in vitro* translation reaction to manufacture tumor-associated or tumor-specific protein.

Extraction of extracellular nucleic acid from blood serum or plasma, and characterization, transcription of complementary RNA, and translation to tumor-associated or tumor-specific protein, provides significant utility, both in the delineation of those who might benefit from therapy, including chemoprevention or assignment of therapy, and in the development of tumor-specific therapies. Sequencing of extracellular nucleic acid or transcription of complementary RNA allows identification or

development of antisense compounds, including synthetic oligonucleotides and other antisense constructs appropriately specific to the extracellular DNA, such as by construction of an expression plasmid such as by adapting the method of Aoki *et al.* (1995, *Cancer Res.* 55: 3810-3816). Similarly, defining tumor characteristics allows

5 identification of specific monoclonal antibody or vaccine therapies appropriately specific to the extracellular DNA. Production of corresponding immunologic protein can be used in the development of tumor-specific monoclonal antibodies. Similarly, translated protein can be used in tumor-specific vaccine development. Furthermore, the extracellular DNA permits a means of defining or allowing the construction of a DNA
10 construct which may be used in vaccine therapy.

Of particular value, the invention allows the development and application of tumor-specific therapies even when only premalignant tumors, early cancers, or occult cancers are present. Thus, the invention allows therapeutic intervention when tumor burden is low, immunologic function is relatively intact, and the patient is not
15 compromised, all increasing the potential for cure.

The invention also provides methods for transcribing RNA complementary to the isolated extracellular nucleic acid from plasma or serum, as well as methods for producing peptides and proteins (or fragments thereof) encoded thereby. Additional methods for using the peptide and proteins as antigens for producing antibodies specific
20 for the peptides and proteins encoded by the extracellular nucleic acids of the invention are also provided. The isolated extracellular nucleic acids of the invention are also used in methods for producing antisense oligonucleotides, either synthetically or using recombinant genetic methods, and the use thereof for affecting gene expression in a cell will be appreciated by one having ordinary skill in the art in view of the methods for
25 isolating and identifying said extracellular nucleic acid provided herein. Vaccine production, as is understood by one with skill in the art, is also enabled using the methods of the invention.

The methods of the invention and preferred uses for the methods of the invention are more fully illustrated in the following Examples. These Examples illustrate certain

aspects of the above-described method and advantageous results. These Examples are shown by way of illustration and not by way of limitation.

5

EXAMPLE 1

Detection of mutated K-ras DNA in the plasma of patients without clinically-diagnosed cancer using an endonuclease-based enrichment method.

Colorectal adenomas are common premalignant neoplasms associated with a risk
10 for development of colorectal cancer. Blood plasma was prospectively obtained from
25 individuals who had K-ras mutated colorectal adenoma. None of the individuals
were known to have cancer, nor did colonoscopy or other testing demonstrate cancer in
any individual. Five to ten milliliters of blood was collected from each individual, and
plasma was fractionated from the whole blood by centrifugation at 400 x g at room
15 temperature for ten minutes. Extracellular DNA was then extracted from the plasma
using a gelatin precipitation method as previously described. The extracted plasma
DNA was amplified with a polymerase chain reaction assay which enriched for mutated
K-ras by employing simultaneous restriction digestion and PCR amplification utilizing
a *Bst*NI restriction endonuclease, the method of which (CARD) is described in full in co-
20 owned and co-pending U.S. Serial No. 08/818,058, the disclosure of which is explicitly
incorporated herein. Mutations in the final digested amplified product were identified
by agarose gel electrophoresis, and further prior to final enzyme digestion by
identification of altered bases using dot blot hybridization. Following amplification, 5 L
of product were applied to a nylon membrane (MSI, Westboro, MA). Replicate blots
25 were hybridized to ³²P-radiolabelled oligonucleotides designed to identify point
mutations in positions 1 and 2 of codons 12 of the K-ras gene (Mutaprobe Human K-ras
12 set, Oncogene Science, Uniondale, N.Y.). Hybridization and wash conditions were
as specified by the membrane's manufacturer. Blots were exposed to x-ray film for 10
hours at -80°C. Mutated K-ras DNA was detected in the plasma of 20 individuals.
30 Sequence analysis demonstrated that the altered base mutation found in the adenoma
was similarly demonstrable in the plasma of the individuals.

This example demonstrates that an enrichment-based assay enables detection of mutated nucleic acid in individuals without clinically-diagnosed cancer; further enables detection of the presence of premalignant tissue including colorectal adenoma; further enables identification of a predictive risk factor; further enables sequence identification of the mutated DNA in blood and further predicts for the sequence of the mutated DNA in tissue.

EXAMPLE 2

10 Detection of mutated P53 DNA in the plasma of individuals without clinically-diagnosed cancer using an enrichment assay.

P53 mutations are common oncogene mutations in both malignant and premalignant tumors. Plasma was obtained prospectively from individuals undergoing colonoscopy and assayed for plasma DNA containing DNA species of P53 mutations at codons 175 and 248. Five to ten milliliters of blood were collected in EDTA-coated siliconized glass vacutainer tubes from each individual assayed. Plasma was fractionated from whole blood by centrifugation at 400 x g at room temperature for ten minutes. Extracellular DNA was then extracted from plasma using a gelatin precipitation extraction as previously described. 7 L of the gelatin-precipitated DNA were then used in an amplification reaction in which P53 DNA was enriched using a restriction endonuclease. Oligonucleotide primers were designed to perform a hemi-nested amplification of a portion of exon 5 for codon 175, wherein the first primer (P53-1) is 5'-GCAGTCACAGCACATGACG-3' (SEQ ID No. 7); the second primer (P53-2) is 5'-AATCAGAGGCCTGGGGAC-3' (SEQ ID No. 8); and the third primer (P53-3) is 5'-GGGCCAGACCTAAGAGCAAT-3' (SEQ ID No. 9). A reaction mixture consisting of 1X *Taq* polymerase (Fisher, Pittsburgh, PA) in a volume of 50 L was prepared. This mixture was amplified by polymerase chain reaction under mineral oil for 20 cycles with denaturation at 94°C for 1 minute, annealing at 58°C for 90 seconds, and extension at 72°C for 90 seconds. This was followed by a CARD step of combined amplification and restriction digestion, in which 5 L of the initial amplification product was transferred to a new reaction mix identical to the K-ras mixture previously described

for CARD in co-pending and co-owned U.S. Serial No. 08/818,058, the disclosure of which is explicitly incorporated herein, except for the use of 10 picomoles each of primers P53-1 and P53-2 in place of K-ras primers, and the substitution of 8 units of restriction enzyme *HinP1* I (New England BioLabs, Beverly, MA) for *Bst*NI. Cycling parameters were as for the CARD K-ras amplification. A final digestion with *HinP1* I was performed prior to identification of mutants by agarose gel electrophoresis. To detect codon 248 P53 mutations a similar CARD assay was performed using primers (P53-4): 5'-TTGGGCCCGGTGTTATCTC-3' (SEQ ID No. 10); and (P53-5): 5'-ATGGTGCGGATGGGCCTC-3' (SEQ ID No. 11), designed to amplify a portion of exon 7. Mutations at codon 248 were enriched and amplified in the same manner as described above for codon 175, except substituting *HinP1* I with *Msp* I (New England BioLabs, Beverly, MA). Mutations in the final digested amplified product were identified by agarose gel electrophoresis.

Extracellular mutated P53 DNA was demonstrated in the plasma of 3 individuals, none of whom had clinically-diagnosed cancer, including one individual having a codon 175 P53 mutation, and 2 individuals having a codon 248 P53 mutation. Colonoscopy demonstrated a premalignant lesion (an adenoma) in each of two of these individuals, and a proliferative disease (a hyperplastic polyp) in the third individual.

This example further demonstrates that extracellular mutated nucleic acid can be detected in the blood of humans without clinically-diagnosed cancer; further that enrichment of the target nucleic acid may be accomplished using a restriction endonuclease; further that mutated P53 DNA may be detected in the blood of humans without clinically-diagnosed cancer; further that mutated P53 DNA may be enriched from the blood of humans without clinically-diagnosed cancer; further that mutated P53 DNA may be detected in the blood of individuals with premalignant neoplasms or proliferative disease; further that P53 DNA provides a predictive risk factor for neoplastic disease. While the example demonstrates detection of P53 mutations at codon 175 and 248, P53 mutations at other codons may similarly be detected in blood by substituting the appropriate primers and restriction enzyme.

EXAMPLE 3

Enhanced identification of humans with premalignant neoplasms using a multiplex assay or assay panel targeting differing mutated oncogenes.

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Since individuals with premalignant neoplasms and conditions may harbor differing mutated oncogenes or differing DNA microsatellite alterations, evaluating blood for a number of differing mutated oncogenes or tumor-associated nucleic acids are thereby more likely to enable identification of those individuals harboring neoplasms, thereby increasing overall sensitivity in identification of said individuals.

10

Plasma was evaluated by a panel of assays which detect mutated K-ras DNA (by the method provided in example 1), and mutated P53 DNA (by the method provided in example 2). Plasma from individuals without clinically-diagnosed cancer shown to harbor colorectal adenoma were evaluated for both extracellular mutated K-ras and mutated P53 DNA. Mutated K-ras but not P53 DNA was demonstrated in the plasma from 20 individuals having K-ras mutated adenoma. However, mutated P53 but not K-ras DNA was additionally demonstrated in the plasma from 2 individuals having wild-type ras adenoma. Together, the panel of both K-ras and P53 assays thereby enabled increased identification of individuals with colorectal adenoma. Increasing the number of mutated oncogenes and tumor-associated nucleic acids detected by the assay array or panel (for example but not limitation by including an APC assay as provided in example 4) thereby increases the ability to identify individuals harboring premalignant neoplasms, thereby increasing the utility of the assay.

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EXAMPLE 4

Detection of an APC gene mutation in plasma from an individual with a colorectal neoplasm.

30

APC gene mutations are common mutations in colorectal premalignant and malignant neoplasms. APC mutations may be demonstrated in blood or tissue by the method as follows: Three overlapping sets of oligonucleotide primers are prepared to the mutation cluster region in exon 15 of the APC gene, which are estimated to account

for over 60% of all APC mutations in colorectal neoplasms. The primer sequences are:

APC-1, 5'-TCCACACCTTCATCTAATGCC-3' (SEQ ID No. 12);

APC-2, 5'-CATTCCACTGCATGGTTCAC-3' (SEQ ID NO. 13);

5 APC-3, 5'-CTGAAGATCCTGTGAGCGAA-3' (SEQ ID No. 14);

APC-4, 5'-TCAGGCTGGATGAACAAGAA-3' (SEQ ID No. 15);

APC-5, 5'-CTTCGCTCACAGGATCTTCA-3' (SEQ ID No. 16);

APC-6, 5'-TTTGAGAGTCGTTGATTGC-3' (SEQ ID No. 17).

10 DNA extracted from plasma, serum, whole blood, or tissue is subjected to polymerase chain reaction amplification with 1 picomole each of either primers APC-1 and APC-2, or primers APC-3 and APC-4 for 25 cycles at a denaturing temperature of 94°C for 60 seconds, annealing at 51°C for 90 seconds, and extension at 72°C for 90 seconds each cycle. The reaction mixture consists of 1X *Taq* buffer, 1.5 mM MgCl₂, 200 micromolar dNTPs, and 1 unit *Taq* polymerase (Fisher, Pittsburgh, PA) in a volume
15 of 50 μ L. Hemi-nested PCR amplification is then performed on 5 μ L of the first amplification product, wherein primers APC-1 and APC-2 are replaced with primers APC-1 and APC-5; and in a separate reaction primers APC-1 and APC-2 replaced with primers APC-2 and APC-3; and in a separate reaction primers APC-3 and APC-4 replaced with primers APC-6 and APC-4. Reaction conditions for the second round of
20 amplification are the same as for the first, except that 10 picomoles of each primer are used, and the cycle number is 35. The PCR products are detected by agarose gel electrophoresis, bands are excised from the gel and DNA isolated using the GeneClean kit (Bio101) according to the manufacturer's instructions. DNA is then asymmetrically reamplified with one primer at 100 picomoles and the other at 2 picomoles for eventual
25 cycle sequencing, wherein primer pairs are: APC-1 and APC-5; APC-2 and APC-3 ; APC-4 and APC-6. The asymmetric amplification is performed as described in McCabe (1990, *PCR Protocols. A guide to methods and applications* (Innis, Gelfand, Sninsky, & White, eds.) Academic Press, pp. 76-83) using approximately 1.4 ng of each PCR product and annealing at 51°C. Cycle sequencing is then performed as described in
30 Brow (1990, *PCR Protocols. A guide to methods and applications* (Innis, Gelfand,

Sninsky, & White, eds.) Academic Press, pp. 189-196) using the limiting primer labeled with ³²P.

5-10 mL of blood was collected in an EDTA-coated siliconized glass vacutainer tube from an individual with colorectal clinically-diagnosed cancer whose tumor was known to harbor an APC mutation consisting of a five base deletion at 3961-3965. Plasma was fractionated from whole blood by centrifugation at 400 x g at room temperature for ten minutes. DNA was then extracted from 200 L of plasma using the High-Pure Viral Nucleic Acid kit (Boehringer Mannheim). The extracted DNA was initially amplified as described above using primers APC-1 and APC-7 (5'-TGCTGGATTGTTCTAGGG-3'; SEQ ID No. 18), then reamplified with primers APC-7 and APC-8 (5'-TCAGACGACACAGGAAGCAG-3'; SEQ ID No. 19) as in the hemi-nested reactions described above. The PCR products were electrophoresed through a 5% agarose gel and examined by ethidium bromide staining. The appropriate region of the gel was excised and prepped by the GeneClean, and this DNA then cycle sequenced as described above, thereby confirming an APC mutation in the plasma DNA.

Plasma from this individual having APC mutated plasma DNA was further evaluated for the presence of mutated K-ras DNA (by the method described in example 1), and mutated P53 DNA (by the method described in example 2). Plasma was negative for both mutated K-ras and mutated P53 DNA in this individual.

This example demonstrates that mutated APC DNA may be detected in blood. Further, detection of mutated APC in blood is indicative of the presence of mutated tissue, and provides a predictive risk factor for neoplastic disease. The example demonstrates that detection of mutated APC in blood is particularly advantageous in predicting for neoplastic disease of a gastrointestinal origin, and particularly a colorectal neoplastic disease.

It will be understood that enriching the extracted DNA for APC by methods as described in the invention will further increase the sensitivity of the assay for detection of mutated APC DNA in blood. It is further shown by this example that it is particularly advantageous to include methods detecting mutated APC within a multiplexed assay or an oncogene or tumor-associated nucleic acid assay panel or array, wherein it is

particularly advantageous to assay blood plasma or serum for mutated APC in addition to mutated K-ras and/or mutated P53. It is well recognized that APC mutations occur early and commonly in premalignant neoplasms such as colorectal adenoma, thus it is particularly evident that it is advantageous to detect mutated APC in the blood of humans without clinically-diagnosed cancer, as to predict for premalignant disease or risk for neoplastic disease, and to further indicate the need for additional diagnostic testing including but not limited to endoscopy, colonoscopy, sigmoidoscopy, radiologic imaging, radionucleotide scanning, ultrasonography, PET scanning, or further evaluation of organ or site specific bodily fluids or stool, wherein bodily fluid includes but is not limited to that collected by drainage, aspiration, direct sampling, and lavage.

EXAMPLE 5

Detection of mutated K-ras DNA in the plasma of a human without clinically-diagnosed cancer having a hyperplastic proliferative condition

Hyperplastic colorectal polyps are non-neoplastic proliferative conditions. Blood was prospectively examined from a human found upon colonoscopy and polypectomy to have a K-ras mutated hyperplastic polyp but not to have cancer. The method of examination was identical to the method provided in example 1. Mutated K-ras DNA was similarly demonstrated in the plasma from the human. This example demonstrates that the methods of the invention enable detection of mutated DNA in the blood of individuals having hyperplastic tissue harboring mutated DNA.

EXAMPLE 6

Detection of mutated K-ras DNA in the plasma of a human with an in-situ (non-invasive) carcinoma

In-situ carcinomas are early cancers which by definition are localized and non-invasive. A human undergoing a colonoscopy was found to have a K-ras

mutated in-situ colorectal carcinoma. Plasma was prospectively obtained from the human and examined using the methods identical to those provided in example 1. Mutated K-ras DNA was similarly detected in the plasma from the human. This example demonstrates that the methods of the invention enable detection of mutated
5 DNA in blood from humans having localized, non-invasive or pre-invasive, in-situ malignancy when the malignancy harbors mutated DNA.

EXAMPLE 7

10 Prophetic Examples of the Use of the Assays of the Invention

The following example is illustrative of potential clinical uses for the assays of the invention.

Case 1

15 A 70 year old woman with a long history of cigarette smoking visits her doctor because she is concerned about her risk of developing lung cancer. A sample of blood is drawn and plasma is prepared. An aliquot of the plasma is incubated with metal beads containing oligonucleotides complementary to exons of the *TP53* tumor suppressor gene bound to their surface. The beads are drawn to the side of the tube with a magnet, and
20 the tube is washed several times with water. The tube is then heated to release the affinity-captured *TP53* DNA fragments, and the solution containing these fragments is applied to a sequencing chip useful for detecting mutations in the *TP53* gene (commercially available from Affymetrix). The machine designed to read the sequencing chip detects a deletion of codon 158 of the *TP53* gene. Confirmation of the presence of
25 the *TP53* mutation is obtained from a sputum sample, and it is concluded that the patient is at high risk for development of lung cancer. She is closely followed by CXR, spiral CT scan, and bronchoscopy, and the patient has blood plasma regularly monitored by a version of the invention adapted specifically for her *TP53* mutation. After enrichment of the plasma for the exon carrying the codon 158 mutation using metal beads followed
30 by amplification, a quantitative assay for the presence of the mutation is used to determine the amount of mutant present in the plasma. For one year, the patient has no

detectable mutant DNA, following which a steady rise in the amount of mutant DNA is noted over six months. No clinical or radiographic evidence of disease is present at this time. The patient begins a P53-directed chemopreventive therapy and after a successful course remains free of disease while being monitored by the codon 158 assay.

5 This example further illustrates the use of the invention in a quantitative fashion, in this case using the affinity-capture version of the invention. A tumor suppressor gene frequently altered in lung cancer (*TP53*) is enriched from the plasma and used as the substrate for a quantitative portion of the invention. This mutant can then be followed and intervention begun early, when chemopreventive therapy is most effective.

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 This clinical vignette is meant as an example of the uses to which the invention may be put, and is not meant in any way to be limitations upon the range or type of assays or extracellular mutant DNAs detectable in the plasma or serum by the invention.

15 It should be understood that the foregoing disclosure emphasizes certain specific embodiments of the invention and that all modifications or alternatives equivalent thereto are within the spirit and scope of the invention as set forth in the appended claims.

What is claimed is:

1. A method for detecting extracellular mutated oncogene DNA in blood from a human or animal without clinically-diagnosed cancer, the method comprising the
5 steps of:

- a) purifying extracellular nucleic acid from blood from a human or animal without clinically-diagnosed cancer to prepare extracted nucleic acid containing a mutated oncogene DNA or a mutated oncogene DNA fragment; and concurrently or sequentially
- 10 b) enriching for the mutated oncogene DNA or a fragment thereof, wherein the mutated oncogene DNA or a fragment thereof is concentrated and/or isolated from the remaining extracted nucleic acid; and
- c) amplifying the enriched mutated oncogene DNA or a fragment thereof, or alternatively amplifying a signal from the enriched mutated oncogene
15 DNA or a fragment thereof; and
- d) detecting the product of the amplified mutated oncogene DNA or the product of its amplified fragment, or the amplified signal of the mutated oncogene DNA or the amplified signal of a fragment thereof.

20 2. A method according to Claim 1 wherein the nucleic acid is purified by heating blood serum or plasma for about 1 to about 10 minutes at a temperature from about 90 degrees C to about 100 degrees C.

25 3. A method according to Claim 1 wherein detection of the product of the amplified mutated oncogene DNA is performed using a detection method selected from the group consisting of gel electrophoresis, single strand conformation polymorphism, heteroduplex analysis, denaturing gradient gel electrophoresis, mismatch cleavage assay, immunological detection methods, nucleic acid hybridization, Southern blot analysis, electrochemiluminescence, reverse dot blot detection, and high-performance liquid
30 chromatography.

4. The method of claim 1 wherein extracellular nucleic acid is purified from the plasma or serum fraction of blood.

5. The method of Claim 1 wherein the enriched mutated oncogene DNA or a fragment thereof of subpart (b) is amplified in subpart (c) using an amplification method selected from the group consisting of polymerase chain reaction, ligase chain reaction, boomerang DNA amplification, Q-beta replication, transcription-based amplification, isothermal nucleic acid sequence based amplification, self-sustained sequence replication assay, strand displacement activation, and cycling probe technology.

6. A method of Claim 1 wherein mutated oncogene DNA or a fragment thereof is enriched in subpart (b) through an endonuclease-mediated restriction digestion.

7. A method of Claim 1 wherein mutated oncogene DNA or a fragment thereof is enriched in subpart (b) by hybridization of the mutated oncogene DNA or a fragment thereof to an oligonucleotide to form a hybridized complex.

8. A method according to Claim 7 wherein the hybridized complex is further subjected to a magnetic field.

9. A method according to Claim 7 wherein the hybridized complex is further subjected to an electric field.

10. A method according to Claim 6 wherein the endonuclease is *Bst*NI, *Hin*P1 I, or *Msp* I.

11. A method for determining the presence of non-hemopoietic cells or tissue having a mutated oncogene allele in a human without clinically-diagnosed cancer, the method comprising the steps of:

- 5 a) purifying extracellular nucleic acid from blood from a human without clinically-diagnosed cancer to prepare extracted nucleic acid containing a mutated oncogene DNA or a mutated oncogene DNA fragment; and concurrently or sequentially
- b) enriching for the mutated oncogene DNA or a fragment thereof, wherein the mutated oncogene DNA or a fragment thereof is concentrated and/or isolated from the remaining extracted nucleic acid; and
- 10 c) amplifying the enriched mutated oncogene DNA or a fragment thereof, or alternatively amplifying a signal from the enriched mutated oncogene DNA or a fragment thereof; and
- d) detecting the product of the amplified mutated oncogene DNA or the product of its amplified fragment, or the amplified signal of the mutated oncogene DNA or the amplified signal of a fragment thereof; and
- 15 e) determining the presence of non-hemopoietic cells or tissue having a mutated oncogene allele in the human thereby, wherein detection of the amplified product or the amplified signal in (d) is indicative of the presence of non-hemopoietic cells or tissue having a mutated oncogene allele in the human.
- 20 12. A method according to Claim 11 wherein detection of the product of the amplified mutated oncogene DNA is performed using a detection method selected from the group consisting of gel electrophoresis, single strand conformation polymorphism, heteroduplex analysis, denaturing gradient gel electrophoresis, mismatch cleavage assay, immunological detection methods, nucleic acid hybridization, Southern blot analysis,
- 25 electrochemiluminescence, reverse dot blot detection, and high-performance liquid chromatography.
13. The method of Claim 11 wherein extracellular nucleic acid is purified from the plasma or serum fraction of blood.

14. The method of Claim 11 wherein the enriched mutated oncogene DNA or a fragment thereof of subpart (b) is amplified in subpart (c) using an amplification method selected from the group consisting of polymerase chain reaction, ligase chain reaction, boomerang DNA amplification, Q-beta replication, transcription-based
5 amplification, isothermal nucleic acid sequence based amplification, self-sustained sequence replication assay, strand displacement activation, and cycling probe technology.

15. A method according to Claim 11, wherein the cells or tissue having a
10 mutated oncogene allele are those of an adenoma.

16. A method according to Claim 11, wherein the tissue having a mutated oncogene allele is associated with colorectal adenoma, cervical dysplasia, atypical squamous metaplasia of the lung, bronchial dysplasia, atypical hyperplasia of the breast,
15 prostatic intraepithelial neoplasia, atypical endometrial hyperplasia, dysplastic nevi of the skin, or Barrett's esophagus.

17. A method according to Claim 11, wherein the method further comprises step (f), whereby upon step (e), locating cells or tissue having a mutated oncogene allele
20 within the human using a diagnostic test selected from the group consisting of colonoscopy, sigmoidoscopy, endoscopy, bronchoscopy, radiologic imaging, ultrasonography, radionucleotide imaging, PET scanning, and evaluation of organ specific bodily fluid, stool, or lavage fluid.

25 18. A method according to Claim 11, wherein the mutated oncogene is mutated K-ras.

19. A method according to Claim 11, wherein the mutated oncogene is mutated APC.

20. A method of Claim 11 wherein mutated oncogene DNA or a fragment thereof is enriched in subpart (b) through an endonuclease-mediated restriction digestion.

5 21. A method of Claim 11 wherein mutated oncogene DNA or a fragment thereof is enriched in subpart (b) by hybridization of the mutated oncogene DNA or a fragment thereof to an oligonucleotide to form a hybridized complex.

10 22. A method according to Claim 21 wherein the hybridized complex is further subjected to a magnetic field.

23. A method according to Claim 21 wherein the hybridized complex is further subjected to an electric field.

15 24. A method according to Claim 20 wherein the endonuclease is *Bst*NI, *Hin*P1 I, or *Msp* I.

20 25. A method for determining an acquired predictive risk factor for a non-hematologic disease in a human without clinically-diagnosed cancer, the method comprising the steps of:

- a) purifying extracellular nucleic acid from blood from a human without clinically-diagnosed cancer to prepare extracted nucleic acid containing a mutated DNA or a mutated DNA fragment; and concurrently or sequentially,
- 25 b) enriching for the mutated DNA or a fragment thereof, wherein the mutated DNA or a fragment thereof is concentrated and/or isolated from the remaining extracted nucleic acid; and
- c) amplifying the enriched mutated DNA or a fragment thereof, or alternatively amplifying a signal from the enriched mutated DNA or a
30 fragment thereof; and

- d) detecting the product of the amplified mutated DNA or the product of its amplified fragment, or the amplified signal of the mutated DNA or the amplified signal of a fragment thereof, whereby said detection determines a predictive risk factor for a non-hematologic disease; and
- 5 e) optionally, demonstrating the absence of the mutated DNA in normal cells from the human, whereby the predictive risk factor is shown to be an acquired predictive risk factor.

26. A method according to Claim 25, wherein detection of the product of the amplified mutated oncogene DNA is performed using a detection method selected from the group consisting of gel electrophoresis, single strand conformation polymorphism, heteroduplex analysis, denaturing gradient gel electrophoresis, mismatch cleavage assay, immunological detection methods, nucleic acid hybridization, Southern blot analysis, electrochemiluminescence, reverse dot blot detection, and high-performance liquid chromatography.

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27. The method of Claim 25 wherein extracellular nucleic acid is purified from the plasma or serum fraction of blood.

28. The method of Claim 25 wherein the enriched mutated oncogene DNA or a fragment thereof of subpart (b) is amplified in subpart (c) using an amplification method selected from the group consisting of polymerase chain reaction, ligase chain reaction, boomerang DNA amplification, Q-beta replication, transcription-based amplification, isothermal nucleic acid sequence based amplification, self-sustained sequence replication assay, strand displacement activation, and cycling probe technology.

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29. A method of Claim 25 wherein mutated oncogene DNA or a fragment thereof is enriched in subpart (b) through an endonuclease-mediated restriction digestion.

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30. A method of Claim 25 wherein mutated oncogene DNA or a fragment thereof is enriched in subpart (b) by hybridization of the mutated oncogene DNA or a fragment thereof to an oligonucleotide to form a hybridized complex.

5 31. A method according to Claim 30 wherein the hybridized complex is further subjected to a magnetic field.

32. A method according to Claim 30 wherein the hybridized complex is further subjected to an electric field.

10

33. A method according to Claim 30 wherein the endonuclease is *Bst*NI, *Hin*P1 I, or *Msp* I.

15 34. A method according to Claim 25, wherein the mutated DNA is a mutated oncogene.

35. A method according to Claim 34, wherein the mutated oncogene is mutated K-ras.

20 36. A method according to Claim 34, wherein the mutated oncogene is mutated APC.

37. A method according to Claim 25, wherein the mutated DNA is DNA having a microsatellite alteration.

25

38. A method according to Claim 25, wherein the non-hematologic disease is one of the following diseases or premalignant conditions: colorectal adenoma, cervical dysplasia, atypical squamous metaplasia of the lung, bronchial dysplasia, atypical hyperplasia of the breast, prostatic intraepithelial neoplasia, atypical endometrial
30 hyperplasia, dysplastic nevi of the skin, or Barrett's esophagus.

39. A method for determining an acquired predictive risk factor for a non-hematologic disease in a human without clinically-diagnosed cancer, the method comprising the steps of:

- 5 a) purifying extracellular nucleic acid from blood from a human without clinically-diagnosed cancer to prepare extracted nucleic acid containing a mutated oncogene DNA or a mutated oncogene fragment, and
- b) amplifying the mutated oncogene DNA or a fragment thereof, or alternatively amplifying a signal from the mutated oncogene DNA or a
10 fragment thereof, and
- c) detecting the product of the amplified mutated oncogene DNA or the product of its amplified fragment, or the amplified signal of the mutated
 oncogene DNA or the amplified signal of a fragment thereof, whereby
 said detection determines a predictive risk factor for a non-hematologic
15 disease; and
- d) optionally, demonstrating the absence of the mutated oncogene DNA in normal cells from the human, whereby the predictive risk factor is shown
 to be an acquired predictive risk factor.

20 40. A method for enriching extracellular mutated DNA or a fragment thereof present within the blood of a human without clinically-diagnosed cancer, the method comprising the steps of :

- a) purifying extracellular nucleic acid from blood of a human without
 clinically-diagnosed cancer to prepare nucleic acid containing a mutated
25 DNA or a mutated DNA fragment, and concurrently or sequentially
- b) hybridizing an oligonucleotide to the mutated DNA or a mutated DNA
 fragment to produce a hybridized product of mutated DNA or a fragment
 thereof; and

- c) separating the hybridized product of mutated DNA or a fragment thereof from the remaining non-hybridized nucleic acid, thereby enriching the mutated DNA or a fragment thereof.

5 41. The method of Claim 40 wherein the extracellular nucleic acid is purified from the plasma or serum fraction of blood.

 42. A method according to Claim 40 wherein the oligonucleotide is further bound to a non-biologic surface.

10

 43. A method according to Claim 40 wherein in subpart (c) the hybridized product is subjected to a magnetic field.

 44. A method according to Claim 40 wherein in subpart (c) the hybridized
15 product is subjected to an electric field.

 45. A method according to Claim 40, wherein the mutated DNA is mutated oncogene DNA.

20 46. A method according to Claim 45, wherein the mutated oncogene is mutated K-ras.

 47. A method according to Claim 45, wherein the mutated oncogene is mutated APC.

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 48. A method according to Claim 45, wherein the mutated oncogene is mutated P53.

 49. A method according to Claim 40, wherein the mutated DNA is DNA
30 having a microsatellite alteration.

50. A method of detecting a mutation in the DNA from the blood or a blood fraction of a human having a disease or premalignant condition in whom the presence of said disease or premalignant condition is unrecognized, wherein the mutation is an acquired mutation of known association with the disease or premalignant condition, and wherein the disease or condition is one of the following: colorectal adenoma, cervical dysplasia, atypical squamous metaplasia of the lung, bronchial dysplasia, atypical hyperplasia of the breast, prostatic intraepithelial neoplasia, atypical endometrial hyperplasia, dysplastic nevi of the skin, or Barrett's esophagus; the method comprising the steps of:

- a) purifying extracellular nucleic acid from blood or a blood fraction from a human in whom the presence of a disease or premalignant condition is unrecognized to prepare extracted nucleic acid containing a mutated DNA wherein the mutation is of known association with the disease or premalignant condition; and
- b) amplifying the mutated DNA, or alternatively amplifying a signal from the mutated DNA; and
- c) detecting the product of the amplified mutated DNA or the amplified signal of the mutated DNA.

51. A method according to Claim 50, wherein the evaluation of blood or a blood fraction from a human for said mutation assists in the identification of said disease or premalignant condition.

52. A method according to Claim 50, wherein extracellular nucleic acid is purified from the plasma or serum fraction of blood.

53. A method according to Claim 51, wherein identification of said disease or premalignant condition is further accomplished using a diagnostic test selected from the group consisting of colonoscopy, sigmoidoscopy, endoscopy, bronchoscopy,

radiologic imaging, ultrasonography, radionucleotide imaging, PET scanning, and evaluation of organ specific bodily fluid, stool, or lavage fluid.

54. A method according to Claim 50, wherein the mutated DNA is a mutated
5 oncogene.

55. A method according to Claim 50, wherein the mutated DNA is a DNA
microsatellite alteration.

56. The method of Claim 50, wherein the mutated DNA of subpart (a) is
10 amplified in subpart (b) using an amplification method selected from the group
consisting of polymerase chain reaction, ligase chain reaction, boomerang DNA
amplification, Q-beta replication, transcription-based amplification, isothermal nucleic
acid sequence based amplification, self-sustained sequence replication assay, strand
displacement activation, and cycling probe technology.

15

57. The method of Claim 50, wherein detection of the product of the
amplified mutated DNA is performed using a detection method selected from the group
consisting of gel electrophoresis, single strand conformation polymorphism,
heteroduplex analysis, denaturing gradient gel electrophoresis, mismatch cleavage assay,
20 immunological detection methods, nucleic acid hybridization, Southern blot analysis,
electrochemiluminescence, reverse dot blot detection, and high-performance liquid
chromatography.

58. A method according to Claim 50, wherein prior to amplification in
25 subpart (b), the mutated DNA in subpart (a) is enriched, wherein the mutated DNA is
thereby concentrated and/or isolated from the remaining extracted nucleic acid.

59. The method of detecting one or more mutated oncogene alleles in the
blood or blood fraction of a human without clinically-diagnosed cancer, wherein the

blood or blood fraction is evaluated for more than one differing mutated oncogene allele in a concurrent or sequential fashion.

5 60. A method according to Claim 59, wherein the blood fraction is plasma or serum.

 61. A method according to Claim 59, wherein a mutated oncogene allele is mutated K-ras.

10 62. A method according to Claim 59, wherein a mutated oncogene allele is an acquired or somatically mutated APC allele.

 63. A method according to Claim 59, wherein a mutated oncogene allele is an acquired or somatically mutated P53 allele.
15

 64. A method for detecting extracellular mutated APC in blood or a blood fraction from a human, the method comprising the steps of:

- 20 a) purifying extracellular nucleic acid from blood or a blood fraction from a human to prepare extracted nucleic acid containing mutated APC DNA or a mutated APC DNA fragment; and
- b) amplifying the mutated APC DNA or a fragment thereof; and
- c) detecting the product of the amplified mutated APC DNA or a fragment thereof.

25 65. A method according to Claim 64, wherein extracellular nucleic acid is purified from the plasma or serum fraction of blood.

 66. A method for determining the presence of cells or tissue having an acquired mutated APC allele in a human, the method comprising the steps of:

- a) purifying extracellular nucleic acid from blood or a blood fraction from a human to prepare extracted nucleic acid containing mutated APC DNA or a mutated APC DNA fragment; and
- b) amplifying the mutated APC DNA or a fragment thereof; and
- 5 c) detecting the product of the amplified mutated APC DNA or a fragment thereof;
- d) determining the presence of cells or tissue having a mutated APC allele in the human thereby, wherein detection of the product of the amplified mutated APC DNA or a fragment thereof in (c) is indicative of the presence of cells or tissue having a mutated APC allele in the human; and
- 10 e) optionally, demonstrating the absence of the mutated APC DNA in normal cells from the human, whereby the demonstrated mutated APC allele is shown to be an acquired mutation.

15 67. A method according to Claim 66, wherein extracellular nucleic acid is purified from the plasma or serum fraction of blood.

68. A method according to Claim 66, wherein the cells or tissue having a mutated APC allele are those of a neoplasm.

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(88) Date of publication of the international search report:
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For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: DETECTION OF EXTRACELLULAR TUMOR-ASSOCIATED NUCLEIC ACID IN BLOOD PLASMA OR SERUM

(57) Abstract: This invention relates to detection of specific extracellular nucleic acid in plasma or serum fractions of human or animal blood associated with neoplastic, pre-malignant or proliferative disease. Specifically, the invention relates to detection of nucleic acid derived from mutant oncogenes or other tumor-associated DNA, and to those methods of detecting and monitoring extracellular mutant oncogenes or tumor-associated DNA found in the plasma or serum fraction of blood by using DNA extraction with or without enrichment for mutant DNA.



WO 01/042504 A3

INTERNATIONAL SEARCH REPORT

Inte I Application No
PCT/US 00/32587

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C12Q1/68 C12P19/34 C07H21/04 C07K1/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

BIOSIS, EPO-Internal, WPI Data, PAJ

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 97 34015 A (PENN STATE RES FOUND) 18 September 1997 (1997-09-18)	1-6, 10-20, 24-29, 33-39, 50-68
Y	* whole document *	7-9, 21-23, 30-32, 40-49

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☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

° Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

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Date of the actual completion of the international search

7 June 2002

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 96 15139 A (AMERICAN HEALTH FOUND ;RONAI ZEEV (US)) 23 May 1996 (1996-05-23)	1-6, 10-20, 24-29, 33-39, 50-68
Y	* whole document *	7-9, 21-23, 30-32, 40-49
X	--- SORENSEN G D ET AL: "Soluble normal and mutated DNA sequences from single-copy genes in human blood." CANCER EPIDEMIOLOGY, BIOMARKERS AND PREVENTION, (1994 JAN-FEB) 3 (1) 67-71., XP001064067 cited in the application	1-6, 10-20, 24-29, 33-39, 50-68
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Y	--- US 5 496 699 A (SORENSEN GEORGE D) 5 March 1996 (1996-03-05) column 1, line 10 -column 9, line 15; figures 1A,1B	1-68
Y	--- WO 95 16792 A (STROUN MAURICE ;ANKER PHILIPPE (CH); VASIOUKHIN VALERI (US)) 22 June 1995 (1995-06-22) example 1	1-68
Y	--- KAHN S M ET AL: "RAPID AND SENSITIVE NONRADIOACTIVE DETECTION OF MUTANT K-RAS GENES VIA ENRICHED PCR AMPLIFICATION" ONCOGENE, vol. 6, no. 6, 1991, pages 1079-1084, XP002191761 ISSN: 0950-9232 cited in the application * whole document *	1-68
Y	--- SIDRANSKY D ET AL: "Identification of ras oncogene mutations in the stool of patients with curable colorectal tumors" SCIENCE, AMERICAN ASSOCIATION FOR THE ADVANCEMENT OF SCIENCE,, US, vol. 256, 3 April 1992 (1992-04-03), pages 102-105, XP000605488 ISSN: 0036-8075 * Abstract * page 103, column 2, paragraph 2 -column 3, paragraph 2 --- -/--	1-68

INTERNATIONAL SEARCH REPORT

Int'l Application No

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	<p>ZEHNDER J ET AL: "CROSS-LINKING HYBRIDIZATION ASSAY FOR DIRECT DETECTION OF FACTOR V LEIDEN MUTATION" CLINICAL CHEMISTRY, AMERICAN ASSOCIATION FOR CLINICAL CHEMISTRY, WINSTON, US, vol. 43, no. 9, September 1997 (1997-09), pages 1703-1708, XP002928152 ISSN: 0009-9147 * Materials and Methods *</p>	7-9, 21-23, 30-32, 40-49
Y	<p>WO 98 58081 A (EXACT LAB INC) 23 December 1998 (1998-12-23)</p> <p>page 5, line 12 -page 7, line 11; example 4</p>	7-9, 21-23, 30-32, 40-49
Y	<p>HORNES E ET AL: "MAGNETIC DNA HYBRIDIZATION PROPERTIES OF OLIGONUCLEOTIDE PROBES ATTACHED TO SUPERPARAMAGNETIC BEADS AND THEIR USE IN THE ISOLATION OF POLY(A) MRNA FROM EUKARYOTIC CELLS" GENETIC ANALYSIS, ELSEVIER, AMSTERDAM, NL, vol. 7, no. 6, 1 October 1990 (1990-10-01), pages 145-150, XP000198013 ISSN: 1050-3862 * whole document *</p>	7-9, 21-23, 30-32, 40-49
Y	<p>LANKIEWICZ SILKE ET AL: "Enhanced RACE method using specific enrichment by biotinylated oligonucleotides bound to streptavidin coated magnetic particles." NUCLEIC ACIDS RESEARCH, vol. 25, no. 10, 1997, pages 2037-2038, XP002201590 ISSN: 0305-1048 * whole document *</p>	7-9, 21-23, 30-32, 40-49
Y	<p>ALBRETSSEN C ET AL: "APPLICATIONS OF MAGNETIC BEADS WITH COVALENTLY ATTACHED OLIGONUCLEOTIDES IN HYBRIDIZATION ISOLATION AND DETECTION OF SPECIFIC MEASLES VIRUS MESSENGER RNA FROM A CRUDE CELL LYSATE" ANALYTICAL BIOCHEMISTRY, vol. 189, no. 1, 1990, pages 40-50, XP008004133 ISSN: 0003-2697 * whole document *</p>	7-9, 21-23, 30-32, 40-49

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INTERNATIONAL SEARCH REPORT

Int. Application No
PCT/US 00/32587

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>VAN MANSFELD A.D.M. ET AL.: "PCR-based Approaches for Detection of Mutated ras Genes"</p> <p>PCR METHODS AND APPLICATIONS, vol. 1, pages 211-216, XP001064082</p> <p>Cold Spring Harbor Lab. Press</p> <p>* whole document *</p> <p>-----</p>	1-68

INTERNATIONAL SEARCH REPORT

ational application No.
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Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☐ Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. ☒ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☒ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: claims 1, 11, 25 (all partially); 2-6, 10, 12-20, 24, 26-29, 33-39, and 50-68 (all completely)

Methods for the detection of oncogenic extracellular DNA from plasma or serum comprising the steps of: (a) purification of extracellular nucleic acid from blood; (b) optionally, enrichment for mutant DNA using selective restriction enzyme digestion; (c) amplification and (d) detection of said mutant oncogene.

2. Claims: 1, 11, 25 (all partially); 7-9, 21-23, 30-32, and 40-49 (all completely)

Methods for the detection of oncogenic extracellular DNA from plasma or serum comprising the steps of: (a) purification of extracellular nucleic acid from blood; (b) enrichment for mutant DNA using oligonucleotide probe hybridization; (c) amplification and (d) detection of said mutant oncogene.

INTERNATIONAL SEARCH REPORT

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